

Benefits and Limitations of *Rhizobium*-Legume Symbiosis and Nitrogen Fixation Under Libyan Saline Soil: Mini Review

Mustafa E. Elsharif*

Department of Microbiology, Faculty of Science, Alasmarya Islamic University, Zliten, Libya.

*E-mail: melsharif@asmarya.edu.ly

فرص استخدام وكفاءة الريزوبيم في تثبيت النيتروجين الجوي تكافليا مع البقوليات في الترب الملحية الليبية

مصطفى الهادي الشريف*

قسم الأحياء الدقيقة، كلية العلوم، الجامعة الأسمرية الإسلامية، زليتن، ليبيا.

Received: 6 March 2019; Revised: 28 August 2019; Accepted: 1 December 2019

Abstract

Biological Nitrogen Fixation (BNF) donates to productivity both straight by increasing the production of legumes and indirectly by improving soil fertility. BNF is the cheapest and most environment-friendly procedure in which nitrogen-fixing bacteria (Rhizobia), interact with leguminous plants. Rhizobium is the most well-known species of the rhizobial group that acts as the primary symbiotic fixer of nitrogen, increased farming of legumes is important for the renewal of nutrient-deficient soils and providing required nutrients which important as food for human beings and animals. Some environmental factors such as salinity, which is become one of the severe difficulties in worldwide farming production. Where soil salinity limits the production of both feed and grain legumes.

Keywords: Legume plants, N-fixation bacteria, Salt tolerances, Libyan soil.

الملخص

يساهم تثبيت النيتروجين البيولوجي في الرفع من الإنتاجية الزراعية بشكل مباشر عن طريق زيادة إنتاج البقوليات وبشكل غير مباشر عن طريق تحسين خصوبة التربة. وتثبيت النيتروجين بيولوجيا هو الإجراء الأرخص والأكثر ملاءمة للبيئة، حيث تقوم البكتيريا المثبتة للنيتروجين المثبتة للنيتروجين المتلاكها النيتروجين الجوي الى أمونيا بواسطة أنزيم النتروجينيز نظرا لامتلاكها جينات تسمى Symbiotic Plasmid وهي المسؤولة عن إحداث العدوى في العقد البكتيرية ومن ثم تثبيت النيتروجين ، وكمية النيتروجين المتوفرة بالتثبيت مهمة جدا في زراعة البقوليات وزيادة نموها وانتاجيتها، وهذا التثبيت ضروري ايضا لتحديد خواص التربة وزيادة خصوبتها، وكذلك للمحافظة على النمو والنشاط الميكروبي في التربة، خاصة تلك التي تعاني من نقص في المغذيات كنتيجة للتأثير السلبي لبعض العوامل البيئية مثل زيادة الملوحة، والتي أصبحت واحدة من المشاكل المحددة في كمية الإنتاج الزراعي العالمي والليبي خاصة، حيث تحد ملوحة التربة من توفير أو زيادة في الانتاج الزراعي.

الكلمات الدالة: الريزوبيم، البقوليات، الملوحة، الترب الليبية.

Benefits and Limitations of Rhizobium-Legume Symbiosis.....

1. Introduction

Around 80% of Earth's atmosphere is nitrogen gas (N₂). Entirely all organisms on the planet use the ammonia (NH₃) form of nitrogen to build up very essentials compound in the life as amino acids, proteins, nucleic acids, and other nitrogen-containing components. BNF is the method that changes inactive N2 into biologically suitable NH3 form, this process is the greatest efficient way to quantity the large volumes of nitrogen needed by legumes to produce high-yielding harvests with a in height protein content (Tate, 1995). On the other hand, this amount of nitrogen fixed by the legumes is influenced by several environmental conditions, soil nitrogen and phosphorous, compatibility of the rhizobia in the soil with the legume, and mycorrhiza colonization (Zahran, 1999; and Nwoko and Sanginga, 1999). Worldwide, the consuming of fertilizer-N expanded from 8 to 17 kg.ha⁻¹ of agricultural land in the 15-year period from 1973 to 1988 (Zahran, 1999). The International Fertilizer Association (IFA) as cited in Heffer and Prud'homme (2016), expects that the demand world for N fertilizer would rise by 1.2% per annum among the base year (average of 2013/14 to 2015/16) and 2020/21 to reach 117.1 T_g N. Similarly, world N demand is predictable up by 1.3% p.a., from 101 T_g N in 2010 to 132 T_g N in 2030 (Heffer and Prud'homme, 2016). Where the annual consuming of fertilizer nutrient in terms of (N, P, and K), has increased also from 0.07 million metric tons (MMT) in 1951-52 to more than 25.95 MMT in 2016-17, and per h intake has increased from less than 1 kg in 1951-52 to the level of 130.8 kg nowadays (Bagal et al., 2018). With the current skill for fertilizer production and the unproductive methods employed for fertilizer application, both the economic and ecological prices of fertilizer usage will eventually become expensive (Zahran, 1999). A widespread range of microorganisms that have the capacity to fix nitrogen, but Rhizobia only have a very great proportion on the nitrogen cycle (Tate, 1995; and Indge, 2000). Whatsoever the correct figure, legume symbioses provide at least 70 million tonnes of N per year, approximately half of this coming from the cool and warm temperature regions and the rest deriving from the tropics (Brockwell et al., 1995), not surprising that atmospheric N₂ fixed symbiotically by the relationship between rhizobium species and legumes represents a renewable foundation of N for crop growing (Wani et al.,1995; and Peoples et al.,1995). Standards estimated for several legume yields and pasture types are often imposing, frequently falling in the range of 200 to 300 kg of N/hectare year, and for example efforts of fixed N for alfalfa, red clover, pea, soybean, cowpea, and vetch were estimated to be about 65 to 335 kg of N/hectare.year (Zahran, 1999). Besides, the rhizobium symbiosis with legumes species is of special importance, making 50% of 175 million tons of total biological N fixation yearly worldwide, in other wards the biological processes account for about 60% for their total contribution to the nitrogen input (Ogutcu et al., 2008). Biochemical nitrogen fertilization can considerably increase legume growth, but rhizobial inoculation characterizes as an economical and environmentally friendly alternative (Ogutcu et al., 2008). Whereas the rate of N fixation differs and lay open to the cultivars and compatibility among the inoculated rhizobium strain and the host cultivar.



ISSN (Prínt): 2413-5267 ISSN (Online): 2706-9966

The rhizobium species that making nodules in a hosts are specific only to this species or strain, and thus inoculation with active strains is advised in soils with no or weak bacterial being there. One of the best methods for sustainable agricultural includes the use of beneficial strains for plants, as they are able to promote plant growth by growing endophytically on plants. In recent years, the increasing worldwide worry regarding to the production of sufficient food to maintain the rising human population, thus has been reinforcing the importance of maintainable intensification of vegetable production.

This review attempts to cover how we deal with the soil salinity as an important problem facing farming production in several areas of Libyan agriculture, and limits legume efficiency, mainly when they depend on symbiotic N₂-fixation. Unfortunately, rhizobium and legumes related factors influencing symbiotic performance still undergoing studies and, poorly documented in home. Additionally, this review aims to explain the line for selection of the best rhizobium strain-legume variety mixture for maximum nitrogen fixation and harvest under saline soils.

2. The World of Legumes and Rhizobium

Legumes (Fabaceae or Leguminosae) is the third main angiosperm family, with more than 727 genera and 19,325 species, and include economically significant grain legumes, oilseed crops, shrubs, forage crops, and tropical or subtropical trees (Singh et al., 2007). Fabaceae are the next important food yield after cereals, they are bases of low-cost dietary and more safety vegetable proteins and minerals when compared with animal foodstuffs such as: egg, meat, and fish, and for other commercial applications (Singh et al., 2007; and Olunike, 2014). Africa has a huge array of indigenous legumes e.g. Cajanus cajan, Lens esculenta, Glycine max, Phaseolus lunatus, Vicia faba and Vigna sinensis (Singh et al., 2007), were legumes belonging to the genera Astragalus, Hippocrepis, Hedysarum, Phaseolus, and Medicago are distributed in Northern Africa (Lioi and Giovannetti, 1987; and Mahdhi et al., 2016). Despite the high number of these legume, only few of them have the capability to establish a symbiotic relationship through soil nitrogen-fixing bacteria (Allito et al., 2014; and Sagasti and Marino, 2015), legumes e.g. Clover, Common Bean, and Alfalfa (Mora et al., 2014). It is predictable about 88% of legume species that examined can form nitrogen fixing nodules with rhizobia, being responsible for up to 80% of the BNF that takes place in farming settings (de la Peña and Pueyo, 2012). Despite the fact that inoculating legumes with rhizobia can achieve considerable increases in nodulation, and biomass yield, but the truth is nitrogen fixation process controlled by the rhizobial strain, legume genotype and the environment factors (Zahran, 1999). While several rhizobium spp. normally occurs in soils but frequently fail to produce active nodulation (Jordan, 1982). However, the rhizobium bacteria are able to convert atmospheric N₂ into NH₄ in presence of the enzyme nitrogenase were the effective nodules occur as pink in color, thus due to a protein called leghemoglobin contains Fe and Mo, and it is responsible for binding oxygen to allows Rhizobium bacteria to live and to fix N2. In addition, symbiosis is not only important for legume crops but also in nitrogen cycle, nitrogen



comes from symbiosis process involving leguminous and Rhizobiaceae. The family of Rhizobiaceae presently involves six genera: Rhizobium, Mesorhizobium, Sinorhizobium, Allorhizobium, Bradyrhizobium and Azorhizobium, which are together referred to as Rhizobia (Datta et al., 2015). As cited in Yoon et al. (2010) the rhizobia or genus rhizobium was first proposed by Frank (1889) and its description was corrected by Young et al. (2001). However, it is classified in phylum Proteobacteria, and class Alphaproteobacteria, in order of Rhizobiales, and in family Rhizobiaceae. The genera of rhizobium have the following characteristics; obligate aerobes, gram negative, non-spore forming chemoheterotrophs (require only simple organic compounds) and grown on a selective medium called yeast-mannitol agar (YMA), motile bacterium they can move using special thread-like structures called flagella, their shaped like short rods, 0.5-0.9 µm wide and 1.2-3.0 μm long, grown at 25-30°C and optimum pH is 6-7. Overall, Rhizobium spp. are able to grow in microaerophilic conditions (Somasegaran and Hoben, 1994), and they are stimulated by root exudates, particularly the root exudates of their host legumes. Table (1) below gives some of the most important rhizobium spp. symbioses with their legumes.

Table 1. Rhizobium *spp.* and the cross-inoculation groups of legumes they modulate.

| No | Rhizobium spp. | Legumes spp. examples |
|----|------------------|------------------------------|
| 1 | Rhizobium etli | Bean and Phaseolus |
| 2 | R. galegae | Galega |
| 3 | R. gallicum | Bean and Phaseolus |
| 4 | R. giardinii | Bean |
| 5 | R. huakuii | Astragalus |
| 6 | R. huautlense | Sesbania |
| 7 | R. indigofera | Indigofera |
| 8 | R. leguminosarum | Clovers; Peas and Vetch |
| 9 | R. mongolense | Medicago |
| 10 | R. tropici | Bean, Leucaena and Phaseolus |

3. Soil Salinity Issues

Not surprising that the opening of 21st century is considered and marked by the global scarcity of water resources, environmental pollution and also increased salinization of soil and water (Shrivastava and Kumar, 2015). By 2050, it is evaluated that about 50% of the world's arable earthly will be influenced by salinity (Wang *et al.*, 2003; and Bartels and Sunkar, 2005). At the present, salinity is a major problematic stressful crop production throughout the world: about 20% of urbane land in the world, and 33% of watered terrestrial, are salt-affected and deteriorated (Machado and Serralheiro, 2017). In addition, approximately 40% of the world's terrestrial surface can be considered as having possible salinity difficulties, and the greatest of these areas are confined to the tropics and Mediterranean regions (Monica *et al.*, 2013). Bouksila (2011), recorded that 5-6 million hectares of arable land in the planet is destroyed yearly due to bad farming management and practices. Furthermore, 5% of this arable land was adversely affected by high level of salt, possibly due to bad agricultural management and practices, anywhere saline soils are common in areas of arid or semi-arid weather (Monica *et al.*, 2013). Several environmental conditions (biotic and abiotic) are limiting aspects to the



ISSN (Prínt): 2413-5267 ISSN (Online): 2706-9966

growth and productive of the N₂-fixing bacteria; include the host symbiont compatibility and the physicochemical situations of the soil. As the major serious threat faced by farming in arid and semi-arid areas is salinity, which counts as the most common issues affecting nitrogen fixation and symbiosis activity (Predeepa and Ravindran, 2010; and Monica et al., 2013), where arid and semiarid climates soils are usually have poor fertility; the rising of salinity and costs of chemical fertilizers and the growing environmental concerns. In compliance with USSL (1954), a saline soil is commonly defined as "a soil with an electrical conductivity of the soil solution extract (ECe) of 4 dS/m". However, desertification and limited freshwater resources resulting in decreasing farming yields and a deterioration in farmer livelihoods that often lead to the abandonment of earlier fertile lands and therefore to desertification. This has led to a histrionic reduction in the terrestrial of productive agricultural lands, in which saline conditions may limit the symbiosis in different ways includes: (a) decreasing the cell number (b) affecting survival and propagation of *Rhizobium* spp. in the soil and rhizosphere, (c) preventing the infection process, (d) reducing photosynthesis, (e) creation of non-functional nodules with irregular structure, and (f) degradation of peribacteroid membrane (Predeepa and Ravindran, 2010; and Moussaid et al., 2015). In addition, the salinity can effect on crops by inhibit the plant's uptake significant nutrients, such nitrate and phosphorous, the salt reasons of ion cytotoxicity, and the osmotic stress (Chinnusamy and Zhu, 2004; and Monica et al., 2014), and finally low soil microbial activity (Yan et al., 2015). The basic solvable salts in soils are the cations Na⁺ (sodium), Ca²⁺ (calcium), Mg²⁺ (magnesium) and K⁺ (potassium), and the anions Cl⁻ (chloride), SO₄⁻² (sulfate), HCO₋₃ (bicarbonate), CO₃⁻² (carbonate) and NO₃- (nitrate) (Tavakkoli et al., 2010; and Yan et al., 2015). However, there is a general consensus on the need to address the problem of soil salinity which developed an issue of main ecological and economic importance in several parts of the planet. Where most leguminous are more sensitive to salinity than their rhizobial fellow members, while the symbiosis process is more sensitive than rhizobia itself (Zahran, 1999). Nevertheless, some rhizobium species are found tolerant to salt stress, and promoting plant growth by developing specific devices to manage with salt stress, including the increase of mineral cations (e.g., K⁺) and the intracellular creation of low-molecular-weight organic solutes such as; trehalose, proline, glycine betaine, and ectoines,. The previous compounds guard cells from desiccation and polyamines osmotic stress by improving the conformation of proteins and biological membranes (Mohammad et al., 1991.; Dong et al., 2017; and Santos and Costa 2002).

In fact some legumes are well at fixing nitrogen than others, but there is a general agreement that rhizobia are able to survive with salinity more than the host legume do, in contrast the difference in nodulation and nitrogen fixation frequently arise in a bacteria strainlegume that tolerant and survival under saline conditions. These bacteria are found in soil or in roots, it has been estimated that 1 g of soil may have a community of 10^9 microbes with Rhizobia (Datta *et al.*, 2015). These days' rhizobium *spp*. has been broadly used to increase legume output in fields, by enhance both growth and nutrient uptake over modulation of hormone-linked phenomena in inoculated vegetation (Tate, 1995).



Benefits and Limitations of Rhizobium-Legume Symbiosis.....

In the *Rhizobium*–legume symbiosis, the route of nitrogen fixation is powerfully related to the physiological status of the host plant. Consequently, an efficient rhizobial strain is not predictable to express its full volume for nitrogen fixation, if limiting causes impose limitations on the potency of the host legume. Numerous solutions have been suggested to managing this issue, in the emerging countries, the most hopeful solution for the near future has become the intensification of agricultural production through either increased cropping intensity and/or higher yields by introducing rhizobium-legume symbiosis as ideal solution to improving soil fertility (Mpepereki et al., 2000), and facing salinity in the rehabilitation of arid lands. Therefore, several publications demonstrating that enhancing and inoculation of stress tolerant strains of rhizobium may improve the nodulation and nitrogen fixation capacity of legumes under harassed circumstances. For instance, the capability of legumes to grow and survive in saline conditions is enhanced when they are inoculated with salt tolerant strains of rhizobia (Zou et al., 1995; and Patil et al., 2014). The inoculation of legumes by salt-tolerant strains of R. leguminosarum bv. trifolii and R. meliloti was reported to enhancing nodulation and N content under salt stress up to 1% NaCl (El-Mokadem et al., 1991). A halotolerant strain EFB1 of Rhizobium meliloti was isolated from nodule of a Melilotus plant grown in a salt marsh, and shows their ability to grow at up to 500 mM NaCl (Lloret et al., 1995). Also Elsheikh (1998) have concluded that rhizobia found to be useful and more salt tolerant than their host legumes. Other studies e.g., Shamseldin and Werner in (2005) have isolated a high salt tolerance strains from Egyptian soils called Rhizobium etli, similarly his fellow countrymen Zahran (1999) have reported that Rhizobium meliloti more salt tolerant than their legume hosts, and can grow at 300 to 700 mM NaCl. In addition, three species of salt-tolerant rhizobia named as; Rhizobium etli, Rhizobium gallicum and Sinorhizobium meliloti have been isolated from Tunisian Oasis (Mnasri et al., 2007). Also Douka, et al. (2008) reported the same founding. Likewise, Ben Romdhane et al. (2008) found from their study that nodulation and yield of chickpea enhanced by inoculation with competitive and salt-tolerant rhizobia. Other study in United Arab Emirates have showed that the isolated rhizobia from desert soils were able to grow and successfully nodulate at high salt concentrations (Sharma et al., 2013). Others in Egypt have concluded that R. leguminosarum by. viciae was able to survive at high salt concentrations at field conditions (Belal et al., 2013). In India Mandal (2014) has isolated five strains of Rhizobium trifolii HK, were they found to be resistances to 3% NaCl salt stresses. Moreover, Sobti et al. in (2015) have concluded from their study that the isolated rhizobia from the Algeria desert soils were able to grow and effectively nodulate even at high salt concentrations. Recently, Mahdhi et al. (2016) have found from 57 isolated bacteria from root nodules of two spontaneous legumes Astragalus corrugatus and Hippocrepis areolata were tolerated to 3% NaCl. Even so, the wide range of salinity levels characteristic of ecosystems where different legumes have been found to fix nitrogen better than others, this suggests that some legume species with specific rhizobial strains can be selected. For example: Vicia faba, Phaseolus vulgaris and Glycine max, were found to be more salt tolerant than other tested legumes (Cordovilla et al., 1999). In contrast, regarding tolerance to NaCl, TU in (1981) reported declines in growth of



ISSN (Prínt): 2413-5267 ISSN (Online): 2706-9966

Rhizobium japonicum when NaCl % was increased from 0.2 to 0.87%. Also 30 isolated of root-nodulating bacteria they tested for their tolerance were found unable to grow at 2% NaCl, except one strain grew at 8% NaCl (Mohamed et al., 2000). Also a study by Thrall et al. (2008) have presents that there was sign for reduced strength of the plant–rhizobial mutualism as part of adaptation to salinity. Furthermore, isolates of rhizobia nodulaing from Vigna Unguiculata L. grown in arid area of Libya by Abdelnaby et al. (2015) showed different response to salinity ranging from sensitive, which unable to grow in 1% NaCl to resistant and grow at 2% NaCl or above. Interestingly introducing mycorrhizal to roots of legumes has been described to stimulate both nodulation and N₂ fixation, particularly in soils low in available phosphor where, phosphorus appears to be a very essential for both nodulation and N₂ fixation (Redecker et al., 1997).

4. Libyan Soil and Crops

Geographically, Libya has four different climates: (a) the Coastal Plains run along the Mediterranean Sea and have Mediterranean climate: warm, dry in Summers, and wet, cold in Winters; (a) the Northern Mountains, includes Jabal Nafusah in the West and Jabal al Akhdar in the East. they have the greater rainfall about 500 and 400 mm respectively and temperatures around 20 °C; (c) the center of Libya, where the desert, and (d) the Southern and Western Mountain part which recognized as a little annual rainfall region from 50 to 150 mm (Abagandura and Park, 2016). In general, the climate is characterized as either semi-arid or arid with very limited rainfall, and its volume and distribution varies from year to year where the agriculture in these area is primary dependent on irrigation. However, El-Asswad (1995) reported that the water level was decline and resulting seawater intrusion with the groundwater resources, which become almost unusable because of their high salinity, also he adding, from 1950 to 1990, the seawater interface advanced 1-2 km inland, and salinity increasing significantly from 150 ppm to around 1,000 ppm in the coastal aquifers. In fact, the majority of the Libyan soil is silty loam and very shallow and coarse, with low in organic matter content and water holding capacity, and about 22.9% of Libyan land is covered by salt flats, rock and dunes (Abagandura and Park, 2016). They added, however, that 95% of total land area in Libya is desert, whereas 4% is grassland, suitable for grazing animals, and 1% is forest, and the most agriculturally productive land limited to a strip abutting the Mediterranean Sea.

Salty soils are common in these areas where passage of soluble salts to the sea does not occur since of low rainfall, they are categorized by the occurrence of high levels of neutral salts in the surface layers' resultant from the capillary rise of water when evaporation surpasses precipitation (Abdelmoumen *et al.*, 1999; Rao and Sharma, 1995; and Belal *et al.*, 2013). As a result, most crops on the Libyan soil do not have sufficient soil nitrogen (Abagandura and Park, 2016), and the country now suffering from increases destroying agricultural land, with very large desert and arid climate. In Summer time some harvests are often irrigated with brackish or reused water, and as a consequence, salts gathering along the



Benefits and Limitations of Rhizobium-Legume Symbiosis.....

soil profile particularly if rainfalls in winter season are not sufficient to eliminate them from the rhizosphere (Abagandura and Park, 2016). Nonetheless, legumes are integral part of Libyan farming systems, and to meet with the consumers' requirements, over recent decades, our concern has been increased about the quantity and the quality of the legume production to feed, the constantly increasing human population has forced us to reinforce the importance of sustainable increase in crop productivity. The requirement for good agricultural practices is revitalizing the attention in BNF and Rhizobia-legumes symbiosis, predominantly those involving economically significant legume harvests in terms of nourishment and forage. Because all of that, more attention should be given to the legumes and rhizobia as they have the greatest quantitative influence on the nitrogen cycle and soil fertility. Refers to the above some rhizobium spp. and strains, which are reported as salt tolerant and efficient nitrogen fixers, have been cultivated (e.g. Bala et al., 1990; Zahran, 1999; and Sobti et al., 2015), this could be used as an alternative to achieve the establishment of symbiosis, where the selection of resistant strains is a significant choice, and can exist in Libyan soils. Overall, a very little work has been carried out on the Libyan legume and on salt tolerance strains of rhizobia, nevertheless the selection of salt-tolerant rhizobia from naturally soils is cheaper and more effective, and better able to nodulate the legume host below saline conditions. Interestingly the native rhizobial strains isolated by Keneni et al. (2010) was found more tolerance to higher salt concentrations of 5% NaCl than the exotic rhizobial strains did. Additionally, results from Ogutcu et al. (2008) have indicated that the chickpea was grow and survive in saline conditions when inoculated with Rhizobium leguminosarum by. ciceri strains that isolated from wild chickpeas. Furthermore, Turkan and Demiral in (2009) have concluded that the efforts to upsurge salt tolerance of harvest plants is important to supply maintainable agriculture and it can improve crop productivity. Therefore, addition of organic materials such as vegetable remains or manures as nutrient sources may encourage rhizobia growth and activity, and could be an important strategy to ameliorate saline soils. At the same time, Rhizobia/legumes enhanced soil structure, erosion protection and better biological diversity.

5. Benefits and Limitations

This paper recognizes the role of biological nitrogen fixation as an unpolluted and more cost-effective way to increase Libyan soil fertility and adaptation to saline condition. In detail, the existence of *Rhizobium*- legume symbioses, which are capable to fix appreciable sum of N₂ under arid conditions is attractive. However, these symbioses signify the best source of the "ideal" fertilizer in arid and semi-arid regions in the country which known as having a high level of salinity and low fertility. On the other hand, under saline conditions the response to rhizobium inoculation, was found to be affected by soil textures, from no response to a highly significant response (Elsheikh,1998). Not surprising that under salt stress some plants differs from others in their tolerances, Pea plants (*Pisum sativum*) showed higher levels of nodule mass and nitrogen fixation than faba bean (*Vicia faba*) plants (Cordovilla *et al.*,1999). Therefore, it is very important to promoting rhizobial growth to contribute by selecting high salt-tolerant strains under salt stress, and understanding the fixation process of nitrogen by



legumes, and how they change within the soil, this will help us to meet with the N potential of our crop production and will help minimize the possibility to reducing the soil salinity. Several recent studies have used molecular techniques e.g. (Shamseldin *et al.*, 2008; Zahran *et al.*, 2013; Moghaddam *et al.*, 2018) including; Plasmid profile, DNA extraction, Polymerase chain reaction (PCR): Metabolomics, Proteomics, and Transcriptomics, to define new genes, proteins, and metabolites involved in a nonliving condition in legumes (abiotic factors). As Libya known as a emerging countries should focus on providing adequate food to their people, which requires both increasing yields and maintain soil fertility. In this way, hope this review assists the Libyan researchers in becoming a more active to introduce the superior rhizobium strains, to produce industrial and saleable biological fertilizer, which is still poorly understood, where, the government through Ministry of Agriculture should recognize that this kind of work must be supported to maintain our food supply.

6. Conclusion

From the above, it can be without difficulty concluded that, nitrogen fixation efficiency depends on rhizobium strain, plant host, salt (a biotic) factor, soil and their interaction. The salt tolerant rhizobium-legume relations well have good possibility for improving the fertility of Libyan saline soils, where introducing tolerant salinity crops such as: barley, oat and species of salt-tolerant olive trees is needed. Whereas a successful management of symbiotic association between leguminous vegetation and their bacterial endosymbionts essentials to improves yields in saline areas. While screening and application of effective tolerance rhizobial strains by using genetic engineering as today an important technological tool will lead to improve *Rhizobium-legume* resistance to salt stress. Further investigation should be carried out about the diversity to select salt tolerant strains of rhizobium bacteria. However, we should take in our account that our emphasis is to select the effective *Rhizobium* strain to achieve maximum benefits from the new technology. It is hereby recommended that there is need to understanding the effects of salinity on symbiotic nitrogen fixation other than the survival of the Rhizobium *spp*. Further advanced study by using molecular tools techniques for identification of rhizobium isolates within host leguminous crop are required.

References

- Abagandura G.O., and Park D. (2016). Libyan Agriculture: A Review of Past Efforts, Current Challenges and Future Prospects. *Journal of Natural Sciences Research*, 6(18): 57-67.
- Abdelmoumen H., Filali-Maltouf A., Neyra M., Belabed A., and El Idrissi M. (1999). Effect of high salts concentrations on the growth of rhizobia and responses to added osmotica. *Journal of Applied Microbiology*, 86: 889–898.
- Abdelnaby M., Elnesairy N.N., Mohamed S.H., and Alkhayali Y.A. (2015). Symbiotic and Phenotypic Characteristics of Rhizobia Nodulaing Cowpea (Vigna Unguiculata L. Walp) Grown in Arid Region of Libya (Fezzan). *Journal of Environmental Science and Engineering* B, 44: 227-239.

- Allito B.B., Ewusi-Mensah N., and Alemneh A.A. (2014). Rhizobia Strain and Host-Legume Interaction Effects on Nitrogen Fixation and Yield of Grain Legume: A Review. *Molecular Soil Biology*, 6(4): 1-12.
- Bagal Y.S., Sharma L.K., Kaur G.P., Singh A., and Gupta P. (2018). Trends and Patterns in Fertilizer Consumption: A Case Study. *International Journal of Current Microbiology and Applied Sciences*, 7(4): 480-487.
- Bala N., Sharma P.K., and Lakshminarayana K. (1990). Nodulation and nitrogen fixation by salinity tolerant rhizobia in symbiosis with tree legumes. *Agriculture, Ecosystems and Environment*, 33: 33-46.
- Bartels D., and Sunkar R. (2005). Drought and salt tolerance in plants. *Crit Rev Plant Sci.*, 24(1): 23–58.
- Belal E.B., Hassan, M.M., and El-Ramady H.R. (2013). Phylogenetic and characterization of salt-tolerant rhizobial strain nodulating faba bean plants. *African Journal of Biotechnology*, 12(27): 4324-4337.
- Ben Romdhane S., Aouani M.S., Trabelsi M., De Lajudie P., and Mhamdi R. (2008). Selection of High Nitrogen-Fixing Rhizobia Nodulating Chickpea (Cicer arietinum) for Semi-Arid Tunisia. *Journal of Agronomy and Crop Science*, 194(6): 413-420.
- Bouksila F. (2011). Sustainability of irrigated agriculture under salinity pressure A study in semiarid Tunisia, Lund: Water Resources Engineering, Lund University, Media-Tryck, Lund, Sweden.
- Brockwell J., Bottomley P.J., and Thies J.E. (1995). Manipulation of rhizobia microflora for improving legume productivity and soil fertility: a critical assessment. *Plant Soil.*, 174(1-2): 143–180.
- Chinnusamy V., and Zhu J.K. (2004). Plant salt tolerance. In: *Plant responses to abiotic stess*. Berlin Heidelberg: Springer, Berlin.Germany.
- Cordovilla M.P., Ligero F., and Lluch C. (1999). Effect of salinity on growth, nodulation and nitrogen assimilation in nodules of faba bean (Vicia faba L). *Applied Soil Ecology*, 11(1): 1-7.
- Datta A., Singh R.K., and Tabassum S. (2015). Isolation, Characterization and Growth of Rhizobium Strains under Optimum Conditions for Effective Biofertilizer Production. *Int. J. Pharm. Sci. Rev. Res.*, 32(1):199-208.
- de la Peña T.C., and Pueyo J.J. (2012). Legumes in the reclamation of marginal soils, from cultivar and inoculant selection to transgenic approaches. *Agron. Sustain. Dev.*, 32: 65–91.
- Dong R., Zhang J., Huan H., Bai C., Chen Z., and Liu G. (2017). High Salt Tolerance of a Bradyrhizobium Strain and Its Promotion of the Growth of Stylosanthes guianensis. *International Journal of Molecular Sciences*, 18(8): 1625-1642.
- Douka C., Apostolakis C., and Skarloy V. (2008). Studies of Rhizobium meliloti isolated from salt-Affected soils. *Annals of Applied Biology*, 88(3): 457–460.
- El-Asswad R.M. (1995). Agriculture Prospects and Water Resources in Libya. Ambio., 24: 324-327.

ISSN (Online): 2706-9966

Elsharíf, 2019

- El-Mokadem M.T., Helemish F.A., Abdel-Wahab S.M., and Abou-El-Nour M.M.. (1991). Salt response of clover and alfalfa inoculated with salt tolerant strains of Rhizobium. *Ain Shams Sci. Bull.*, 28(B): 441–468.
- Elsheikh E. (1998). Response of Legume-Rhizobium Symbiosis to Salinity in the Sudan: a Review. *Agric. Sci.*, 6(2): 142-156.
- Heffer P., and Prud'homme M. (2016). Global nitrogen fertilizer demand and supply: trend, current level and outlook. Proceedings of *International Nitrogen Initiative Conference*, "Solutions to improve nitrogen use efficiency for the world", 4–8 December, Melbourne, Australia.
- Indge B. (2000). The Nitrogen Cycle. Biological Sciences Review, 13: 25-27.
- Jordan D.C. (1982). Transfer of Rhizobium japonicum to Bradyrhizobium a genus of slow growing, root nodule bacteria from leguminous Plants. *International Journal of systemic Bacteriology*. 32: 136-139.
- Keneni A., Assefa F., and Prabu P.C. (2010). Characterization of Acid and Salt Tolerant Rhizobial Strains Isolated from Faba Bean Fields of Wollo, Northern Ethiopia. *J. Agr. Sci. Tech.*, 12: 365-376.
- Lioi L., and Giovannetti M. (1987). Variable effectivity of three vesicular-arbuscular mycorrhizal endophytes in Hedysarum coronarium and Medicago sativa. *Biol Fertil Soils*, 4: 193-197.
- Lloret J., Bolanos L., Lucas M., Peart J.M., Brewin J.M., Bonilla. L., and Rivilla R. (2015). Microbiology Ionic Stress and Osmotic Pressure Induce Different Alterations in the Lipopolysaccharide of a Rhizobium meliloti strain. *Applied and Environmental Microbiology*, 61(10): 3701–3704.
- Machado R.M.A., and Serralheiro R.P. (2017). Review Soil Salinity: Effect on Vegetable Crop Growth. Management Practices to Prevent and Mitigate Soil Salinization. *Horticulturae*, 3(30): 1-13.
- Mahdhi M., Houidheg N., Mahmoudi N., Msaadek A.A., Rejili M., and Mars M. (2016). Characterization of Rhizobial Bacteria Nodulating Astragalus corrugatus and Hippocrepis areolata in Tunisian Arid Soils. *Polish Journal of Microbiology*, 65(3): 331–339.
- Mandal H.K. (2014). Isolation of Salt Tolerant Strains of Rhizobium Trifolii. *International Journal of Agriculture and Food Science Technology*, 5(4): 325-332.
- Moghaddam M., Sabzevar A., Zolfaghari M., and Lakzian A. (2018). Phenotypic and molecular characterization of Sinorhizobium meliloti strains isolated from the roots of Medicago sativa in Iran. *Biological Journal of Microorganism*, 6(24): 29-39.
- Mohammad R.M., Akhavan-Kharazian M., Campbell W.F., and Rumbaugh M.D. (1991). Identification of salt and drought tolerant Rhizobium meliloti L. strains. *Plant Soil*, 134: 271–276.
- Mnasri B., Mrabet M., Laguerre G., Aouani M.E., and Mhamd R. (2007). Salt-tolerant rhizobia isolated from a Tunisian oasis that are highly effective for symbiotic N2-fixation with Phaseolus vulgaris constitute a novel biovar (bv. mediterranense) of Sinorhizobium meliloti. *Archives of Microbiology*, 187(1): 79–85.

- Mohamed S.H., Smouni A., Neyra M., Kharchaf D., and Filali-Maltouf A. (2000). Phenotypic characteristics of root-nodulating bacteria isolated from Acacia spp. grown in Libya. *Plant and Soil*. 224 (2): 171–183.
- Monica N., Vidican R., Pop R., and Rotar I. (2013). Stress Factors Affecting Symbiosis Activity and Nitrogen Fixation by Rhizobium Cultured in vitro. *ProEnvironment*, 6: 42-45.
- Monica N., Vidican R., Ioan Rotar I., Stolan V., Rodica R., and Miclea R. (2014). Plant Nutrition Affected by Soil Salinity and Response of Rhizobium Regarding the Nutrients Accumulation. *ProEnvironment*, 7: 71-75.
- Mora Y., Rafael R., Vargas-Lagunas C., Peralta H., Guerrero G., Aguilar A., Encarnación S., Girard L., and Mora J. (2014). Nitrogen-Fixing Rhizobial Strains Isolated from Common Bean Seeds: Phylogeny, Physiology, and Genome Analysis. *Applied and Environmental Microbiology*, 80(18): 5644–5654.
- Moussaid S., Domínguez-Ferreras A., Muñoz S., Aurag J., and Sanjuán J. (2015). Increased trehalose biosynthesis improves Mesorhizobium ciceri growth and symbiosis establishment in saline conditions. *Symbiosis.*, 67: 103–111.
- Mpepereki S., Javeheri F., Davis P., Giller K.E., and Vance C.P. (2000). Soybean and Sustainable agriculture. Promiscuous soybeans in Southern Africa. *Fields Crop Research*, 65: 137-149.
- Nwoko H., and Sanginga N. (1999). Dependence of promiscuous soybeans and herbaceous legumes on the arbuscular myhcorrhiza fungi and their response to Bradyrhizobium inoculation in low P soils. *Applied Soil Ecology*, 13: 131-143.
- Turkan I., and Demiral T. (2009). Recent developments in understanding salinity tolerance. *Environ*. *Exp. Bot.*, 67: 2–6.
- Ogutcu H., Algur O.F., Elkoca E., and Kantar F. (2008). The Determination of Symbiotic Effectiveness of Rhizobium Strains Isolated from Wild Chickpeas Collected from High Altitudes in Erzurum. *Turkish Journal of Agriculture and Forestry*, 32: 241-248.
- Olunike A. (2014). Utilization of Legumes in the Tropics. *Journal of Biology, Agriculture and Healthcare*, 4(12): 77-84.
- Patil S.M., Patil D.B., Patil M.S., Gaikwad P.V., Bhamburdekar S.B., and Patil P.J. (2014). Isolation, characterization and salt tolerance activity of Rhizobium sp. from root nodules of some legumes. *Int. J. Curr. Microbiol. App. Sci.*, 3(5): 1005-1008.
- Peoples M.B., Herridge D.F., and Ladha J.K. (1995). Biological nitrogen fixation: an efficient source of nitrogen for sustainable agricultural production. *Plant Soil*, 174: 3–28.
- Predeepa R.J., and Ravindran D.A. (2010). Nodule formation, distribution and symbiotic efficacy of Vigna unguiculata L. under different soil salinity regimes. *Emir. J. Food Agric.*, 22(4): 275-284.
- Rao D.L.N., and Sharma P.C. (1995). Effectiveness of rhizobial strains for chickpea under salinity stress and recovery of nodulation on desalinization. *Indian J. Exp. Biol.*, 33: 500-504.

- Redecker D., von Berswordt-Wallrabe P., Beck D.P., and Werner D. (1997). Influence of inoculation with arbuscular mycorrhizal fungi on stable isotopes of nitrogen in Phaseolus vulgaris. *Biol. Fertil. Soils*, 24: 344–346.
- Sagasti M.T., and Marino D. (2015). PGPR sand nitrogen fixing legumes: a perfect team for efficient Cd phytoremediation? *Plant Biotechnology*, 81(6): 1-9.
- Santos H., and da Costa M.S. (2002). Compatible solutes of organisms that live in hot saline environments. *Environ. Microbiol.*, 4: 501–509.
- Shamseldin A., and Werner D. (2005). High salt and high pH tolerance of new isolated Rhizobium etli strains from Egyptian soils. *Curr Microbiol.*,50(1): 11-16.
- Shamseldin A., Sadowsky M., El-Saadani M., and Sun An C. (2008). Molecular Biodiversity and Identification of Free Living Rhizobium Strains from Diverse Egyptian Soils as Assessed by Direct Isolation Without Trap Hosts. *American-Eurasian J. Agric. & Environ. Sci.*, 4(5): 541-549.
- Sharma S.R., Rao N.K., Gokhale T.S., and Ismail S. (2013). Isolation and characterization of salt-tolerant rhizobia native to the desert soils of United Arab Emirates. *Emir. J. Food Agric.*, 25(2): 102-108.
- Shrivastava P., and Kumar R. (2015). Review, Soil salinity: A serious environmental issue and plant growth promoting bacteria as one of the tools for its alleviation. *Saudi Journal of Biological Sciences*. 22: 123–131.
- Singh R.J., Chung G.H., and Nelson R.L. (2007). Landmark research in legumes. *Genome*, 50: 525–537.
- Sobti S., Belhadj H.A., and Djaghoubi A. (2015). Isolation and Characterization of The Native Rhizobia Under Hyper-Salt Edaphic Conditions in Ouargla (southeast Algeria). *Energy Procedia*, 74: 1434–1439.
- Somasegaran P., and Hoben H.J. (1994). *Handbook for Rhizobia: Methods in Legumes-Rhizobium Technology*. Springer-Verlag Inc., New York.
- Tate R.L. (1995). Soil microbiology (symbiotic nitrogen fixation). John Wiley & Sons, Inc., New York.
- Tavakkoli E., Rengasamy P., and McDonald G.K. (2010). High concentrations of Na⁺ and Cl⁻ ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. *Journal of Experimental Botany*, 61(15): 4449–4459.
- Thrall P.H., Bever J.D., and Slattery J.F. (2008). Rhizobial mediation of Acacia adaptation to soil salinity: evidence of underlying trade-offs and tests of expected patterns. *Journal of Ecology*, 96: 746–755.
- Tu J.C. (1981). Effect of salinity on Rhizobium root hair interaction, nodulation and growth of Soybean. *Can. J. Plant. Sci.*, 61: 231-239.
- USSL (1954). Salinity Laboratory Staff, Diagnosis and Improvement of Saline and Alkali Soils. In: USDA Agric. Handbook No. 60. U.S Government Printing Office, Washington, D.C.

JOURNAL OF MARINE SCIENCES & ENVIRONMENTAL TECHNOLOGIES Vol. 5, Issue No. 2 (December-2019)



Benefits and Limitations of Rhizobium-Legume Symbiosis.....

- Wang W., Vinocur B., and Altman A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta*, 218(1): 1–14.
- Wani S.P., Rupela O.P., and Lee K.K. (1995). Sustainable agriculture in the semi-arid tropics through biological nitrogen fixation in grain legumes. *Plant Soil*, 174: 29–49.
- Yan N., Marschnerc P., Caoa W., Zuoa C., and Qin W. (2015). Influence of salinity and water content on soil microorganisms. *International Soil and Water Conservation Research*, 3: 316–323.
- Yoon J.H., Kang So-Jung., Hoon J., Yi H., Oh T., and Choong-Min. R. (2010). Rhizobium soli sp. nov., isolated from soil. *International Journal of Systematic and Evolutionary Microbiology*. 60: 1387–1393.
- Zahran H.H. (1999). Rhizobium-Legume Symbiosis and Nitrogen Fixation under Severe Conditions and in an Arid Climate. *Microbiology and Molecular Biology Reviews*, 63(4): 968–989.
- Zahran H.H., Chahboune R., Moreno S., Bedmar E.J., Abdel-Fattah M., Yasser M.M., and Mahmoud A.M. (2013). Identification of rhizobial strains nodulating Egyptian grain legumes. *International Microbiology*, 16: 157-163.
- Zou N., Dart P.J., and Marcar N. (1995). Interaction of salinity and rhizobial strain on growth and N₂ fixation by Acacia ampliceps. *Soil. Biol. Biochem.* 27: 409-413.